Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis

The International Multiple Sclerosis Genetics Consortium* & the Wellcome Trust Case Control Consortium 2*

Multiple sclerosis is a common disease of the central nervous system in which the interplay between inflammatory and neurodegenerative processes typically results in intermittent neurological disturbance followed by progressive accumulation of disability¹. Epidemiological studies have shown that genetic factors are primarily responsible for the substantially increased frequency of the disease seen in the relatives of affected individuals^{2,3}, and systematic attempts to identify linkage in multiplex families have confirmed that variation within the major histocompatibility complex (MHC) exerts the greatest individual effect on risk⁴. Modestly powered genome-wide association studies (GWAS)⁵⁻¹⁰ have enabled more than 20 additional risk loci to be identified and have shown that multiple variants exerting modest individual effects have a key role in disease susceptibility¹¹. Most of the genetic architecture underlying susceptibility to the disease remains to be defined and is anticipated to require the analysis of sample sizes that are beyond the numbers currently available to individual research groups. In a collaborative GWAS involving 9,772 cases of European descent collected by 23 research groups working in 15 different countries, we have replicated almost all of the previously suggested associations and identified at least a further 29 novel susceptibility loci. Within the MHC we have refined the identity of the HLA-DRB1 risk alleles and confirmed that variation in the HLA-A gene underlies the independent protective effect attributable to the class I region. Immunologically relevant genes are significantly overrepresented among those mapping close to the identified loci and particularly implicate T-helper-cell differentiation in the pathogenesis of multiple sclerosis.

We performed a large GWAS as part of the Wellcome Trust Case Control Consortium 2 (WTCCC2) project. Cases were recruited through the International Multiple Sclerosis Genetics Consortium (IMSGC) and compared with the WTCCC2 common control set^{12,13} supplemented by data from the control arms of existing GWAS. We introduced a number of novel quality control methods for processing these data sets (see Supplementary Information), which ultimately provided reliable information from 9,772 cases and 17,376 controls (Fig. 1a). After single nucleotide polymorphism (SNP)-based quality controls, data from 465,434 autosomal SNPs, common to all internally and externally generated data sets, were available for analysis.

The multi-population nature of our study (Fig. 1a, b) afforded an opportunity to assess various published approaches for controlling the potential confounding effects of population structure, several of which (in the event) proved unhelpful (see Supplementary Information). Although not common in primary GWAS undertaken to date, the challenge of combining data across populations, in contexts where not all case samples have controls available from the same population (thus precluding standard meta-analytical techniques), may become more routine as study sizes increase.

We attempted analyses of the non-United Kingdom (UK) data with the now widespread technique of using principal components as covariates to correct for structure. However, even use of all seven top principal components that captured genome-wide effects in our data resulted in an unacceptably high genomic inflation: for example, the genomic control factor¹⁴ (λ) was $\lambda = 1.2$. We tried to reduce the genomic inflation by discarding the case samples that seemed least well matched to control sets. Removal of half the available cases in this fashion only reduced λ to 1.1. In another approach to handling structure, statistical clustering algorithms were successful in identifying subgroups of the data within which cases and controls seemed well matched for ancestry (see Supplementary Fig. 17). However, tests within these subgroups combined via fixed-effects meta-analysis also yielded unacceptably high genomic inflation ($\lambda > 1.4$) in an analysis with seven matched subgroups of cases and controls. Lastly, we applied a novel variance components method (similar to one described previously¹⁵), separately to the UK and non-UK data sets, which explicitly accounts for correlations among the



Figure 1 | **Distribution of cases and controls. a, b**, All cases and controls were drawn from populations with European ancestry; cases from 15 countries and controls from 8. **a**, Numbers of case (red) and control (black) samples from each country. **b**, The projection of samples onto the first two principal components of genetic variation, with cases shown on the left and controls on the right. The axes are orientated to approximate the geography, and samples are colour coded as indicated in the legend. NZ, New Zealand. We genotyped the cases (9,772) and some Swedish controls (527) using the Illumina Human 660-Quad platform, and the UK controls (5,175, the WTCCC2 common control set^{12,13}) using the Illumina 1.2M platform. All other controls were genotyped externally using various Illumina genotyping systems (see Supplementary Information).

*A list of authors and their affiliations appears at the end of the paper; membership of both consortia is listed in Supplementary Information.

phenotypes of individuals resulting from relatedness, allowing us to deal successfully with all sources of structure in our samples (see Supplementary Information for details of the linear mixed model we used). For example, the genomic inflation was reduced to $\lambda = 0.995$ in the UK and 1.016 in the non-UK data (see also Supplementary Information). After fixed-effects meta-analysis of the results from the UK and non-UK data sets, the inflation factor was $\lambda = 1.045$. We adopted this approach for all subsequent non-MHC association analyses.

Outside the MHC we identified 95 distinct regions having at least one SNP associated with multiple sclerosis at $P_{\rm GWAS} < 1 \times 10^{-4.5}$; in six of these 95 regions conditional analysis revealed an additional SNP showing association to the same locus (one locus containing two such SNPs). In total we took all 102 SNPs forward to replication, which we performed using data from previously reported multiple sclerosis GWAS^{8,9} and the iControl database (excluding any WTCCC controls previously used in these studies). In total, the replication analysis included data from 4,218 cases and 7,296 controls. These were considered in six independent strata after which results were combined through a fixed-effects meta-analysis. For 98 of the 102 SNPs, the same allele was overrepresented in cases compared to controls. Twenty three of the 26 previously known or strongly suggested multiple-sclerosisassociated loci were replicated in our primary GWAS with $P_{\rm GWAS} < 1 \times 10^{-3}$. Our GWAS and replication also revealed another 29 novel associated regions (defined as having $P_{\rm GWAS} < 1 \times 10^{-4.5}$, one-sided $P_{\rm replication} < 0.05$ and $P_{\rm combined} < 5 \times 10^{-8}$), and a further 5 regions with strong evidence for association (with $P_{\rm GWAS} < 1 \times 10^{-4.5}$, one-sided $P_{\rm replication} < 0.05$ and $P_{\rm combined} < 5 \times 10^{-7}$). In one previously reported locus and two novel loci, additional SNPs were identified as being conditionally important in explaining risk. Just over one third of the identified loci overlap with regions already confirmed as associated with at least one other autoimmune disease (according to the GWAS catalogue, http://www.genome.gov/gwastudies/). Results for both the previously established and novel loci are shown in Fig. 2 and Supplementary Tables 1–3; and details of all 102 SNPs taken to replication are available in Supplementary Data.

To assess objectively the collective evidence across the associated regions for particular classes of genes, we performed statistical analyses to look for enrichment of genes with similar function. We first identified



Figure 2 | Regions of the genome showing association to multiple sclerosis. Columns from left to right: first, evidence for association from the linear mixed model analysis of the discovery data (thresholded at - $\log_{10}P$ value = 12). Non-MHC regions containing associated SNPs are shown in red and are labelled with the rs number (bold for newly identified loci, black for strong evidence, grey for previously reported) and risk allele of the most significant SNP. Asterisk indicates that the locus contains a secondary SNP signal. Second, OR and 95% confidence intervals estimated from the meta-analysis of the discovery and replication data (+ indicates estimates for previously known loci from discovery data only). Third, risk allele frequency estimates in each of the control populations used in the study (each is shown as a vertical bar on a scale from 0 to 1 going left to right). For each region of association the number of genes is reported (fifth), and where non-zero a candidate gene is given (fourth). Black dots indicate that the candidate gene is physically the nearest gene (sixth) included in the 'immune system process' GO term (seventh). Eight, when the mostsignificant SNP tags an SNP predicted to have an impact on the function of the candidate gene this is indicated. Where such an SNP exists, the gene involved is selected as the candidate gene; otherwise the nearest gene is selected unless there are strong biological reasons for a different choice. The final column indicates SNPs that are correlated ($r^2 > 0.1$) with SNPs reported to be associated with other autoimmune (AI) diseases. CeD, coeliac disease; CrD, Crohn's disease; PS, psoriasis; RA, rheumatoid arthritis; T1D, type 1 diabetes; UC, ulcerative colitis. An interactive version of the figure is available at http://www.well.ox.ac.uk/wtccc2/ms.

the nearest gene to the lead SNP in each of the (52) regions of association and used the Gene Ontology (GO) database¹⁶ to define sets of functionally related genes (GO terms). We then tested whether the set of nearest genes was enriched for particular GO terms using Fisher's exact test. The GO terms having the most significant enrichment include genes linked to lymphocyte function ($P = 3.2 \times 10^{-11}$, odds ratio (OR) = 35.96) and in particular those with a role in T-cell activation and proliferation ($P = 1.85 \times 10^{-9}$, OR = 40.85). These are representative of a larger group associated with various components of the GO 'immune system process' ($P = 8.6 \times 10^{-11}$, OR = 9.12). A similar analysis based on all genes in or near association regions showed similar enrichment, as did independent analyses based on nearest gene or all genes in our next tier of signals, the 42 regions taken to replication but not meeting the thresholds above for association (see Supplementary Data). Although GO immune system genes only account for 7% of human genes, in 30% of our association regions the nearest gene to the lead SNP is an immune system gene. As an illustration, Fig. 3 shows a schematic of genes involved in the T-helper-cell differentiation pathway; a notable number show strong evidence for association with multiple sclerosis, particularly those acting as cell surface receptors. We infer from this pathway analysis of our GWAS signals that specific classes of immune system genes are especially important in the pathogenesis of multiple sclerosis.

Our screen not only implicates a multitude of genes coding for cytokine pathway (*CXCR5, IL2RA, IL7R, IL7, IL12RB1, IL22RA2, IL12A, IL12B, IRF8, TNFRSF1A, TNFRSF14, TNFSF14*), co-stimulatory (*CD37, CD40, CD58, CD80, CD86, CLECL1*) and signal transduction (*CBLB, GPR65, MALT1, RGS1, STAT3, TAGAP, TYK2*) molecules of immunological relevance, but also relates to previously reported environmental risk factors such as vitamin D^{9,17} (*CYP27B1, CYP24A1*) and therapies for multiple sclerosis including natalizumab¹⁸ (*VCAM1*) and daclizumab¹⁹ (*IL2RA*). There is a relative absence of genes relevant to potential pathways for neurodegeneration independent of inflammation (*GALC, KIF21B*).

To refine our understanding of the MHC associations in multiple sclerosis we imputed classical human leukocyte antigen (HLA) types at six loci (*A*, *B*, *C*, *DQA1*, *DQB1* and *DRB1*)²⁰ and analysed these alongside the SNPs (see Supplementary Information for validation; at alleles responsible for the major signals described later, estimated specificity was at least 0.99 and sensitivity was at least 0.98, except for DRB1*13:03, where it was 0.88). Primary discovery was focused on the UK cohort with candidate signals being validated through support from additional case–control cohorts. Because of the extensive linkage disequilibrium within the MHC, we identified associated alleles in a stepwise



manner, selecting the most strongly associated to include in a general model, in turn, if $P_{\rm UK} < 10^{-4}$ and $P_{\rm combined} < 10^{-9}$ (Supplementary Information). At each stage we explored possible interactions and departures from the simple model in which risk increases multiplicatively with each additional copy of the relevant allele (additive increase on the log-odds scale) within the logistic risk framework.

Using this approach we found that DRB1*15:01 has the strongest association with multiple sclerosis among all classical and SNP alleles, with a consistent effect between cohorts ($P < 1 \times 10^{-320}$; Fig. 4a). The data are consistent with an additive effect on the log-odds scale for each additional allele. Conditioning on DRB1*15:01, we confirmed the presence of a protective class I allele and identified the signal as being driven by HLA-A*02:01 (as previously suggested²¹), with a consistent effect size across cohorts ($P = 9.1 \times 10^{-23}$; Fig. 4a). Again, we found no strong evidence for departure from additivity on the log-odds scale or statistical interaction with DRB1*15:01. Conditioning on both DRB1*15:01 and A*02:01 revealed additional risk associated with the strongly linked alleles DRB1*03:01 and DQB1*02:01 ($P = 3.6 \times 10^{-10}$; Fig. 4a; note that we cannot separate these alleles but for simplicity refer only to DRB1*03:01 later). Further conditioning identified an additional *DRB1* risk allele DRB1*13:03 ($P = 1.3 \times 10^{-11}$; Fig. 4a). Although no other classical alleles meet the above criteria, we did observe several SNPs providing independent signals, the strongest coming from rs9277535_G (combined OR 1.28, $P = 2.2 \times 10^{-22}$), an allele known to be in linkage disequilibrium with DPB1*03:01 ($r^2 = 0.37$)²².

Analysis of the MHC SNP data using a genealogical method (GENECLUSTER)²³ offers an alternative means of relating our results to classical HLA alleles that provides additional insight into the underlying genetic architecture (see Supplementary Information). Figure 4b shows genealogical trees relating the classical alleles at DRB1 and HLA-A, together with the estimated evolutionary position of the mutations predicted by GENECLUSTER, as most completely modelling the association. At HLA-DRB1, three mutations are predicted, each of which implicates a clade of haplotypes carrying particular DRB1 alleles. All of the DRB1 alleles we have shown to be independently associated are included in these clades, each corresponding to a particular mutation. In addition, the analysis also explains why those haplotypes carrying the *08:01 allele have previously been shown to increase risk^{24,25} as they carry the same mutation as those bearing *13:03. At HLA-A, the predicted protective mutation is also concordant with our regression analysis of classical alleles in implicating *02:01 but, in addition, predicts that *68:01, *02:05 and *02:06 carry the same protective allele. All of these secondary predictions (increased risk from DRB1*08:01 and protection from HLA-A*68:01, *02:05 and *02:06) are supported in our regression

> Figure 3 Graphic representation of the T-helper-cell differentiation pathway. The figure is derived from an image generated by Ingenuity Pathway Analysis (IPA) software version 8.8 (Ingenuity Systems). Alphanumeric labels indicate the individual genes and gene complexes (nodes) included in the pathway (note that some are included more than once). Coloured nodes are those containing a gene implicated by proximity to an SNP showing evidence of association. Red, in bold or grey in Fig. 2 (plus MHC class II region and TNF); orange, other loci in Fig. 2 or discovery P value $< 1 \times 10^{-4.5}$ and consistent replication data; yellow, discovery *P* value $< 1 \times 10^{-3}$. Other molecules (proteins, vitamins etc) may also be of relevance in these processes but are not included here as they are not currently listed as being part of this particular pathway in the IPA database.

LETTER RESEARCH



Figure 4 | **Results for the main MHC alleles. a**, Forest plots for each of the primary HLA alleles (HLA-A*02:01, DRB1*15:01, DRB1*03:01 and DRB1*13:03) showing consistency of effect across the populations and combined OR of 0.73, 3.1, 1.26 and 2.4, respectively (whiskers indicate 95% confidence intervals). **b**, The genealogical trees estimated for *DRB1* (top) and *HLA-A* (bottom). These trees were constructed using classical HLA and SNP typing data available from the HapMap CEU haplotype data. Each left-hand branch of the tree terminates on a set of haplotypes carrying a particular HLA allele. The coloured dots indicate the mostly likely locations for a disease-associated mutation as predicted by the GENECLUSTER program²³. In the

analysis of classical alleles but the power to detect them in the primary analyses is limited because each allele occurs at very low frequency.

We found no evidence for genetic associations with clinical course, severity of disease or month of birth, and no evidence of interaction with gender or DRB1*15:01 in any part of the genome (see Supplementary Information). However, analysis with respect to age at onset replicated the previously suggested association with the DRB1*15:01 allele²⁶. Although no other part of the genome contained individual SNPs showing strong evidence for association, risk alleles determining susceptibility are collectively more closely associated with age at onset than expected by chance, indicating that individual genetic susceptibility is inversely correlated with age at onset.

Our GWAS—large for any complex trait having a prevalence of 1:1,000 and involving diverse populations of European descent—has identified 29 novel susceptibility loci. Four mutations, one from class I and three from class II, with effects modelled in a simple multiplicative manner within and across loci are sufficient to account for most of the risk attributable to the MHC (see Supplementary Information). Although our data do not address the issue of which components within the nervous system are initially damaged by the inflammatory response, the overrepresentation of genes that influence T-cell maturation provides independent and compelling evidence that the critical disease mechanisms primarily involve immune dysregulation.

More generally, our study reinforces the view that the GWAS design, combined with very large experimental sample sizes and careful statistical analysis, provides valuable insights into the genetic architecture of common complex diseases. Here, this approach has identified many associated genetic variants close to genes, which are both individually interesting and collectively illuminate the roles of key biological pathways. It also provides indirect evidence that many more common variants of small effect contribute to genetic susceptibility for multiple sclerosis. Simple models, in which the previously known and newly identified variants affect risk multiplicatively, both within and across loci, explain a meaningful proportion (\sim 20%, see Supplementary Information) of genetic risk for the disease. Important challenges lie ahead in understanding overlap between the genetic basis for

DRB1 tree, the blue dot captures a risk effect attributable to all haplotypes carrying the *15:01 allele. The green dot captures a risk effect carried by all haplotypes carrying the *03:01 allele and the red dot captures a risk effect on haplotypes carrying *13:03 or *08:01. In the *HLA-A* plot, the orange dot is a protective mutation lying at the root of all *02:01, *02:05, *02:06 and *68:01 alleles. The blue dot in brackets denotes a branch containing those *03:01 haplotypes that also carry DRB1*15:01; the GENECLUSTER prediction here is thus a reflection, due to linkage disequilibrium of the risk attributable to DRB1*15:01. The terminal branches are labelled with the allele carried by the haplotype and its frequency.

susceptibility in the context of different autoimmune diseases, and in uncovering the functional mechanisms underlying these associations.

METHODS SUMMARY

Details of case ascertainment, processing and genotyping, together with sample and genotyping quality control are provided in Supplementary Information. Statistical methods developed for testing the reliability of externally generated data sets, detecting samples with non-European ancestry, correcting for structure, classical HLA imputation and meta-analysis are also outlined in Supplementary Information. Results for all scans and all reported loci are described in detail in Supplementary Information.

Received 4 February; accepted 2 June 2011.

- 1. Compston, A. & Coles, A. Multiple sclerosis. Lancet 372, 1502–1517 (2008).
- Dyment, D. A., Yee, I. M., Ebers, G. C. & Sadovnick, A. D. Multiple sclerosis in stepsiblings: recurrence risk and ascertainment. *J. Neurol. Neurosurg. Psychiatry* 77, 258–259 (2006).
- Hemminki, K., Li, X., Sundquist, J., Hillert, J. & Sundquist, K. Risk for multiple sclerosis in relatives and spouses of patients diagnosed with autoimmune and related conditions. *Neurogenetics* 10, 5–11 (2009).
- The International Multiple Sclerosis Genetics Consortium. A high-density screen for linkage in multiple sclerosis. Am. J. Hum. Genet. 77, 454–467 (2005).
- 5. The International Multiple Sclerosis Genetics Consortium. Risk alleles for multiple sclerosis identified by a genomewide study. *N. Engl. J. Med.* **357**, 851–862 (2007).
- The Wellcome Trust Case Control Consortium & The Australo-Anglo-American Spondylitis Consortium. Association scan of 14,500 nonsynonymous SNPs in four diseases identifies autoimmunity variants. *Nature Genet.* **39**, 1329–1337 (2007).
 Baranzini, S. E. *et al.* Genome-wide association analysis of susceptibility and
- Baraizini, S. E. et al. Genome-wide association analysis of susceptibility and clinical phenotype in multiple sclerosis. *Hum. Mol. Genet.* 18, 767–778 (2009).
- De Jager, P. L. et al. Meta-analysis of genome scans and replication identify CD6, IRF8 and TNFRSF1A as new multiple sclerosis susceptibility loci. Nature Genet. 41, 776–782 (2009).
- The ANZgene Consortium. Genome-wide association study identifies new multiple sclerosis susceptibility loci on chromosomes 12 and 20. *Nature Genet.* 41, 824–828 (2009).
- Sanna, S. et al. Variants within the immunoregulatory CBLB gene are associated with multiple sclerosis. Nature Genet. 42, 495–497 (2010).
- The International Multiple Sclerosis Genetics Consortium (IMSGC). Evidence for polygenic susceptibility to multiple sclerosis—the shape of things to come. Am. J. Hum. Genet. 86, 621–625 (2010).
- The U.K. Parkinson's Disease Consortium & the Wellcome Trust Case Control Consortium 2. Dissection of the genetics of Parkinson's disease identifies an

additional association 5' of SNCA and multiple associated haplotypes at 17q21. *Hum. Mol. Genet.* **20**, 345–353 (2011).

- The Genetic Analysis of Psoriasis Consortium & the Wellcome Trust Case Control Consortium 2. A genome-wide association study identifies new psoriasis susceptibility loci and an interaction between *HLA-C* and *ERAP1*. *Nature Genet.* 42, 985–990 (2010).
- Devlin, B. & Roeder, K. Genomic control for association studies. *Biometrics* 55, 997–1004 (1999).
- 15. Kang, H. M. *et al.* Variance component model to account for sample structure in genome-wide association studies. *Nature Genet.* **42**, 348–354 (2010).
- Ashburner, M. et al. Gene Ontology: tool for the unification of biology. Nature Genet. 25, 25–29 (2000).
- 17. Pierrot-Deseilligny, C. & Souberbielle, J. C. Is hypovitaminosis D one of the
- environmental risk factors for multiple sclerosis? Brain 133, 1869–1888 (2010).
 18. Steinman, L. A molecular trio in relapse and remission in multiple sclerosis. Nature Rev. Immunol. 9, 440–447 (2009).
- Bielekova, B. et al. Effect of anti-CD25 antibody daclizumab in the inhibition of inflammation and stabilization of disease progression in multiple sclerosis. Arch. Neurol. 66, 483–489 (2009).
- Leslie, S., Donnelly, P. & McVean, G. A statistical method for predicting classical HLA alleles from SNP data. Am. J. Hum. Genet. 82, 48–56 (2008).
- Brynedal, B. et al. HLA-A confers an HLA-DRB1 independent influence on the risk of multiple sclerosis. PLoS ONE 2, e664 (2007).
- Field, J. et al. A polymorphism in the HLA-DPB1 gene is associated with susceptibility to multiple sclerosis. PLoS ONE 5, e13454 (2010).
- Su, Z., Cardin, N., Donnelly, P. & Marchini, J. A Bayesian method for detecting and characterizing allelic heterogeneity and boosting signals in genome-wide association studies. *Stat. Sci.* 24, 430–450 (2009).
- Barcellos, L. F. et al. Heterogeneity at the HLA-DRB1 locus and risk for multiple sclerosis. Hum. Mol. Genet. 15, 2813–2824 (2006).
- Dyment, D. A. et al. Complex interactions among MHC haplotypes in multiple sclerosis: susceptibility and resistance. *Hum. Mol. Genet.* 14, 2019–2026 (2005).
- sclerosis: susceptibility and resistance. *Hum. Mol. Genet.* 14, 2019–2026 (2005).
 Masterman, T. *et al.* HLA-DR15 is associated with lower age at onset in multiple sclerosis. *Ann. Neurol.* 48, 211–219 (2000).

Supplementary Information is linked to the online version of the paper at www.nature.com/nature.

Acknowledgements The principal funding for this study was provided by the Wellcome Trust (085475/B/08/Z, 085475/Z/08/Z, 075491/Z/04/Z and 068545/Z/02). The work was also supported by National Institutes of Health (Al076544, NS032830, NS049477, NS19142, NS049510, NS26799, NS43559, NS067305, CA104021, RR020092, RR024992 and K23N/S048869), US National Multiple Sclerosis Society (RG 4201-A-1), Nancy Davis Foundation, Cambridge NIHR Biomedical Research Centre, UK Medical Research Council (G0700061, G0000934), Multiple Sclerosis Society of Great Britain and Northern Ireland (898/08), Wolfson Royal Society Merit Award, Peter Doherty fellowship, Lagrange Fellowship, Harry Weaver Neuroscience Scholarships, Australian National Health and Medical Research Council (NHMRC), Australian Research Council Linkage Program Grant, JHH Charitable Trust Fund, Multiple Sclerosis Research Australia, Health Research Council New Zealand, National MS Society of New Zealand, Wetenschappelijk Onderzoek Multiple Sclerose, Bayer Chair on Fundamental Genetic Research regarding the Neuroimmunological Aspects of Multiple Sclerosis, Biogen Idec Chair Translational Research in Multiple Sclerosis, FWO-Vlaanderen, Belgian Neurological Society, Danish Multiple Sclerosis Society, Neuropromise EU grant (LSHM-CT-2005-018637), Center of Excellence for Disease Genetics of the Academy of Finland, Sigrid Juselius Foundation, Helsinki University Central Hospital Research Foundation, Bundesministerium für Bildung und Technologie (KKNMS consortium Control MS), Deutsche Forschungsgemeinschaft, Institut National de la Santé et de la Recherche Médicale (INSERM), Association pour la Recherche sur la Sclérose En Plaques (ARSEP), Association Française contre les Myopathies (AFM), Italian Foundation for Multiple Sclerosis (FISM grants 2002/R/40, 2005/R/10, 2008/R/11 and 2008/R/15), Italian Ministry of Health (grant Giovani Ricercatori 2007 - D.Igs 502/92), Regione Piemonte (grants 2003, 2004, 2008, 2009), CRT Foundation, Turin, Moorfields/UCL Institute of Ophthalmology NIHR Biomedical Research Centre, Norwegian MS Register and Biobank, Research Council of Norway, South-Eastern and Western Norway regional Health Authories, Ullevål University Hospital Scientific Advisory Council, Haukeland University Hospital, Amici Centro Sclerosi Multipla del San Raffaele (ACESM), Association of British Neurologists, Spanish Ministry of Health (FISPI060117), Bibbi and Niels Jensens Foundation, Montel Williams foundation, Hjärnfonden and Swedish medical research council (8691), Stockholm County Council (562183), Swedish Council for Working life and Social Research, Gemeinnützige Hertie Stiftung, Northern California Kaiser Permanente members and Polpharma Foundation, and Washington University Institute of Clinical and Translational Sciences-Brain, Behavioral and Performance Unit. We acknowledge use of data from the British 1958 Birth Cohort, the UK National Blood Service, the popgen biobank, the KORA and MONICA Augsburg studies, the Accelerated Cure Project, the Brigham & Women's Hospital PhenoGenetic Project, the Swedish CAD project, the Norwegian Bone Marrow Donor Registry, the Children's Hospital of Philadelphia (CHOP), the Swedish Breast Cancer study, BRC-REFGENSEP (Pitié-Salpêtrière Centre d'Investigation Clinique (CIC) and Généthon) and HYPERGENES (HEALTH-F4-2007-201550). Projects received support from the German Ministry of Education and Research, the Helmholtz Zentrum München-National Research Center, the German National Genome Research Network (NGFN), the LMUinnovativ, the Knut and Alice Wallenberg Foundation, the Center for Applied Genomics from the Children's Hospital of Philadelphia Development Award, the Agency for Science & Technology and Research of Singapore, and the Susan G. Komen Breast Cancer Foundation. We thank S. Bertrand, J. Bryant, S. L. Clark, L. Collimedaglia, G. Coniglio, J. S. Conquer, B. Colombo, T. Dibling, G. Eckstein, J. C. Eldred, G. Fischer, S. Gamble, P. Gregersen, R. Guerrero, C. Hind, P. Lichtner, L. Moiola, H. Mousavi, R. Naismith, R. J. Parks, R. Pearson, V. Pilato, M. Radaelli, E. Scarpini, C. R. Stribling, T. Strom, S. Taylor, D. Vukcevic and A. Wilk for their help and support. Detailed acknowledgements are available in Supplementary Information. This manuscript is dedicated to the memory of L. Peltonen, a member of both the IMSGC and the WTCCC2, in recognition of her contributions to, and her leadership in, human genetics.

Author Contributions Details of individual contributions are listed in Supplementary Information.

Author Information Reprints and permissions information is available at www.nature.com/reprints. The authors declare no competing financial interests. Readers are welcome to comment on the online version of this article at www.nature.com/nature. Correspondence and requests for materials should be addressed to A.C. (alastair.compston@medschl.cam.ac.uk; for the IMSGC) or P.D. (donnelly@well.ox.ac.uk; for the WTCC2).

The International Multiple Sclerosis Consortium & the Wellcome Trust Case Control Consortium 2 (Membership of both consortia is listed in Supplementary Information.)

Consortium 2 (Membership of both consortia is listed in Supplementary Information.) Stephen Sawcer^{1*}, Garrett Hellenthal^{2*}, Matti Pirinen^{2*}, Chris C. A. Spencer^{2*}, Nikolaos A. Patsopoulos^{3,4,5}, Loukas Moutsianas⁶, Alexander Dilthey⁶, Zhan Su², Colin Freeman², Sarah E. Hunt⁷, Sarah Edkins⁷, Emma Gray⁷, David R. Booth⁸, Simon C. Potter⁷, An Goris⁹, Gavin Band², Annette Bang Oturai¹⁰, Amy Strange², Janna Saarela¹¹, Céline Bellenguez², Bertrand Fontaine¹², Matthew Gillman⁷, Bernhard Hemmer¹³, Rhian Gwilliam⁷, Frauke Zipp^{14,15}, Alagurevathi Jayakumar⁷, Roland Martin¹⁶, Stephen Leslie¹⁷, Stanley Hawkins¹⁸, Eleni Giannoulatou², Sandra D'alfonso¹⁹, Hannah Blackburn⁷, Filippo Martinelli Boneschi²⁰, Jennifer Liddle⁷, Hanne F. Harbo^{21,22}, Marc L. Perez⁷, Anne Spurkland²³, Matthew J. Waller⁷, Marcin P. Mycko²⁴, Michelle Ricketts⁷, Manuel Comabella²⁵, Naomi Hammond⁷, Ingrid Kockum²⁶, Owen T. McCann⁷, Maria Ban¹, Pamela Whittaker⁷, Anu Kemppinen¹, Paul Weston⁷, Clive Hawkins²⁷, Sara Widaa⁷, John Zajicek²⁸, Serge Dronov⁷, Neil Robertson²⁹, Suzannah J. Bumpstead⁷, Lisa F. Barcellos^{30,31}, Rathi Ravindrarajah⁷, Roby Abraham²⁷, Lars Alfredsson³², Kristin Ardlie⁴, Cristin Aubin⁴, Amie Baker¹, Katharine Baker²⁹, Sergio E. Baranzini³³, Laura Bergamaschi¹⁹, Roberto Bergamaschi³⁴, Allan Bernstein³¹, Achim Berthele¹³, Mike Boggild³⁵, Jonathan P. Bradfield³⁶, David Brassat³⁷, Simon A. Broadley³⁸, Dorothea Buck¹³, Helmut Butzkueven^{39,40,41,42}, Ruggero Capra⁴³, William M. Carroll⁴⁴, Paola Cavalla⁴⁵, Elisabeth G. Celius²¹, Sabine Cepok¹³, Rosetta Chiavacci³⁶, Françoise Clerget-Darpoux⁴⁶, Katleen Clysters⁹, Giancarlo Comi²⁰, Mark Cossburn²⁹, Isabelle Cournu-Rebeix¹², Mathew B. Cox⁴⁷, Wendy Cozen⁴⁸, Bruce A. C. Cree³³, Anne H. Cross⁴⁹, Daniele Cusi⁵⁰, Mark J. Daly^{4,51,52}, Emma Davis⁵³, Paul I. W. de Bakker^{3,4,54,55}, Marc Debouverie⁵⁶, Marie Beatrice D'hoogh Joseph Glessner³⁶, Refujia Gomez³³, Olivier Gouf⁴², Colin Graham⁶⁵, Struan F. A. Grant^{36,66,67}, Franca Rosa Guerini⁶⁸, Hakon Hakonarson^{36,66,67}, Per Hall⁶⁹, Anders Hamsten⁷⁰, Hans-Peter Hartung⁷¹, Rob N. Heard⁸, Simon Heath⁷², Jeremy Hobart²⁸, Muna Hoshi¹³, Carmen Infante-Duarte⁷³, Gillian Ingram²⁹, Wendy Ingram²⁸, Talat Islam⁴⁴, Maja Jagodic²⁶, Michael Kabesch⁷⁴, Allan G. Kermode⁴⁴, Trevor J. Kilpatrick^{39,40,75}, Cecilia Kim³⁶, Norman Klopp⁷⁶, Keijo Koivisto⁷⁷, Malin Larsson⁷⁰, Mark Lathrop⁷², Jeannette S. Lechner-Scott^{47,78}, Maurizio A. Leone⁷⁹, Virpi Leppä^{11,80}, Ulrika Liljedahl⁸¹, Izaura Lima Bomfim²⁶, Robin R. Lincoln³³, Jenny Link²⁶, Jianjun Liu⁸², Áslaug R. Lorentzen^{22,83}, Sara Lupoli^{50,84}, Fabio Macciardi^{50,85}, Thomas Mack⁴⁸, Mark Marriott^{39,40}, Vittorio Martinelli²⁰, Deborah Mason⁸⁶, Jacob L. McCauley⁸⁷, Frank Mentch³⁶, Inger-Lise Mero^{21,83}, Tania Mihalova²⁷, Xavier Montalban²⁵, John Mottershead^{88,89}, Kjell-Morten Myhr^{90,91}, Paola Naldi⁷⁹, William Ollier⁵³, Alison Page⁹², Aarro Palotic^{7,11,93,94}, Jean Pelletier⁹⁵, Laura Piccio⁴⁹, Trevor Pickersgill²⁹, Fredrik Piehl²⁶, Susan Pobywajlo⁵, Hong L. Quach³⁰, Patricia P. Ramsay³⁰, Mauri Reunane⁹⁶, Richard Reynolds⁹⁷, John D. Rioux⁹⁸, Mariaemma Rodegher²⁰, Sabine Roesner¹⁶, Justin P. Rubio³⁹, Ina-Maria Rücker⁷⁶, Marco Salvetti⁹⁹, Erika Salvi^{50,100}, Adam Santaniello³³, Catherine A. Schaefer³¹, Stefan Schreiber^{58,101}, Christian Schulze¹⁰², Rodney J. Scott⁴⁷, Finn Sellebjerg¹⁰, Krzysztof W. Selmaj²⁴, David Sexton¹⁰³, Ling Shen³¹, Brigid Simms-Acuna³¹, Sheila Skidmora¹, Patrick M. A. Sleiman^{36,66}, Cathrine Smestad²¹, Per Soelberg Sørensen¹⁰, Helle Bach Søndergaard¹⁰, Jim Stankovich⁶¹, Richard C. Strange²⁷, Ana-Maja Sulonen^{11,80}, Emilie Sundqvist²⁶, Ann-Christine Syväne⁸¹, Francesca Taddeo¹⁰⁰, Bruce Taylo⁶¹, Jenefer M. Blackwell^{104,105}, Pentit Tienari¹⁰⁶, Elvira Bramon¹⁰⁷, Ayman To Alastair Compston¹*

¹University of Cambridge, Department of Clinical Neurosciences, Addenbrooke's Hospital, BOX 165, Hills Road, Cambridge CB2 0QQ, UK. ²Wellcome Trust Centre for Human Genetics, Roosevelt Drive, Oxford OX3 7BN, UK. ³Division of Genetics, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston,

Massachusetts 02115, USA. ⁴Broad Institute of Harvard University and Massachusetts Institute of Technology, Cambridge, 02142 Massachusetts, USA. ⁵Center for Neurologic Diseases, Department of Neurology, Brigham and Women's Hospital, Boston, Massachusetts 02115, USA. ⁶Department of Statistics, University of Oxford, Oxford OX1 3TG, UK. ⁷Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge CB10 1SA, UK. ⁸Westmead Millennium Institute, University of Sydney, New South Wales 2145, Australia. ⁹Laboratory for Neuroimmunology, Section of Experimental Neurology, Katholieke Universiteit Leuven, 3000 Leuven, Belgium. ¹⁰Danish Multiple Sclerosis Center, Department of Neurology, Copenhagen University Hospital, Rigshospitalet, 2100 Copenhagen, Denmark.¹¹Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki 00290, Finland.¹²INSERM UMR S 975 CRICM, UPMC, Département de neurologie Pitié-Salpêtrière, AP-HP, 75013 Paris, France. ¹³Department of Neurology, Klinikum Rechts der Isar, Technische Universität München, Ismaninger Strasse 22, 81675 Munich, Germany. ¹⁴Department of Neurology, University Medicine Mainz, Johannes Gutenberg University Mainz, Langenbeckstr. 1, 55131 Mainz, Germany. ¹⁵Max Delbrueck Center for Molecular Medicine, Robert-Rössle-Str. 10, 13092 Berlin, Germany. ¹⁶Institute for Neuroimmunology and Clinical MS Research (inims), ¹⁷Department of Clinical Pharmacology, Falkenried 94, D-20251 Hamburg, Germany.
¹⁷Department of Clinical Pharmacology, University of Oxford, Old Road Campus Research Building, Old Road Campus, Oxford OX3 7DQ, UK.¹⁸Queen's University Belfast, University Road, Belfast BT7 1NN, Northern Ireland, UK. 19 Department of Medical Sciences and Interdisciplinary Research Center of Autoimmune Diseases (IRCAD), University of Eastern Piedmont, 28100 Novara, Italy. ²⁰Department of Neurology, Institute of Experimental Neurology (INSPE), Division of Neuroscience, San Raffaele Scientific Institute, Via Olgettina 58, 20132 Milan, Italy. ²¹Department of Neurology, Oslo University Hospital, N-0407 Oslo, Norway. ²²Department of Neurology, University of Oslo, N-0318 Oslo, Norway. ²³Institute of Basal Medical Sciences, University of Oslo, N-0317 Oslo, Norway. ²⁴Department of Neurology, Laboratory of Neuroimmunology, Medical University of Lodz, Kopcinskiego 22, 90-153 Lodz, Poland. ²⁵Clinical Neuroimmunology Unit, Multiple Sclerosis Center of Catalonia (CEM-Cat), Vall d'Hebron University Hospital, Barcelona 08035, Spain. ²⁶Department of Clinical Neurosciences, Centre for Molecular Medicine CMM, L8:04, Karolinska Institutet, Karolinska University Hospital, SE-171 76 Stockholm, Sweden. ²⁷Keele University Medical School, Stoke-on-Trent ST4 7NY, UK. ²⁸Peninsula College of Medicine and Dentistry, Universities of Exeter and Plymouth, Clinical Neurology Research Group, Tamar Science Park, Plymouth PL6 8BX, UK. ²⁹Department of Neurology, University Hospital of Wales, Heath Park, Cardiff CF14 4XW, UK. ³⁰Genetic Epidemiology and Genomics Laboratory, Division of Epidemiology, School of Public Health, University of California, Berkeley, California 94720-7356, USA. ³¹Kaiser Permanente Northern California Division of Research, 2000 Broadway, Oakland, California 94612, USA. ³²Institute of Environmental Medicine, Karolinska Institutet, Box 210, 171 77 Stockholm, Sweden. ³³Department of Neurology, University of California San Francisco, 505 Parnassus Avenue, S-256, San Francisco, California 94143-0435, USA. ³⁴Neurological Institute C. Mondino, IRCCS, 27100 Pavia, Italy. ³⁵The Walton Centre for Neurology and Neurosurgery, Liverpool L7 9LJ, UK. ³⁶Center for Applied Genomics, The Children's Hospital of Philadelphia, 3615 Civic Center Blvd., Philadelphia, Pennsylvania Children's Hospital of Philadelphila, 3615 Civic Center Bivd., Philadelphila, Pennsylvania 19104, USA.³⁷INSERM U 1043 et Pôle Neurosciences, Hopital Purpan, 31059 Toulouse, France. ³⁸School of Medicine, Griffith University, 4222, Australia.³⁹Florey Neuroscience Institutes, University of Melbourne, Victoria 3010, Australia.⁴⁰Royal Melbourne Hospital, Parkville, Victoria 3050, Australia.⁴¹Box Hill Hospital, Monash University, Box Hill 3128, Australia. ⁴²Department of Medicine, RMH Cluster, University of Melbourne, Victoria 3010, Australia. ⁴³Multiple Sclerosis Centre, Department of Neurology, Ospedali Civili di Brescia, 25018 Brescia, Italy. ⁴⁴Centre for Neuromuscular and Neurological Disorders, University of Western Australia, Perth, Western Australia 6009, Australia. ⁴⁵Department of Neurosciences, University of Turin, A.O.U. San Giovanni Battista, 10126 Turin, Italy. ⁴⁶INSERM U669, Univ Paris-Sud, 94800 Villejuif, France. ⁴⁷University of Newcastle, University Drive, Callaghan, New South Wales 2308, Australia. ⁴⁸Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Norris Comprehensive Cancer Center, 1441 Eastlake Ave. Room 4453, Los Angeles, California 90033, USA. ⁴⁹Department of Neurology, Washington University, St Louis, Missouri 63110, USA. ⁵⁰University of Milan, Department of Medicine, Surgery and Dentistry, AO San Paolo, University of Milan, c/o Filarete Foundation, Viale Ortles 22/4, 20139 Milano, Italy.⁵¹Harvard Medical School, Boston, Massachusetts 02115, USA.⁵²Center for Human Genetic Research, Massachusetts General Hospital, Boston, Massachusetts 02114, USA. ⁵³The UK DNA Banking Network, Centre for Integrated Genomic Medical Research, University of Manchester M13 9PT, UK. ⁵⁴Department of Medical Genetics, Division of University of Manchester M13 9PT, UK. ⁵⁴Department of Medical Automated Theorem 2010 (2010) (2010 University of Manchester M13 9P1, UK. ⁵⁷ Department of Medical Genetics, Division of Biomedical Genetics, University Medical Center Utrecht, 3508 GA, Utrecht, The Netherlands. ⁵⁵ Julius Center for Health Sciences and Primary Care, University Medical Center Utrecht, 3508 GA, Utrecht, The Netherlands. ⁵⁶Service de Neurologie, Hôpital Central, 54035 Nancy, France. ⁵⁷ National Multiple Sclerosis Center, 1820 Melsbroek, Belgium. ⁵⁸Institute for Clinical Molecular Biology, Christian-Albrechts-University, Kiel, D-24105, Germany. ⁵⁹Department of Neurology, Tampere University Hospital, FIN-33014 Tampere, Finland. ⁶⁰University of Tampere, Medical School, Tampere 33014, Finland. ⁶¹Menzies Research Institute Tasmania University of Tasmania Private Rag 23 Finland. ⁶¹Menzies Research Institute Tasmania, University of Tasmania, Private Bag 23, Hobart, Tasmania 7000, Australia.⁶²Department of Neurological Sciences, Centro Dino Ferrari, University of Milan, Fondazione Cà Granda, Ospedale Maggiore Policlinico, 20122 Milan, Italy. 63 Centro Studi Sclerosi Multipla, Ospedale di Gallarate, 21013 Gallarate (VA), Italy. ⁶⁴Service de Neurologie, Fondation Ophtalmologique Adolphe de Rothschild, 75019 Paris, France. ⁶⁵Belfast Health and Social Care Trust, City Hospital, Belfast BT9 7AB, Northern Ireland, UK. ⁶⁶Division of Genetics, The Children's Hospital of Philadelphia, 3615 Civic Center Blvd., Philadelphia, Pennsylvania 19104, USA. ⁶⁷Department of Pediatrics, University of Pennsylvania School of Medicine, 3615 Civic Center Blvd., Philadelphia, Pennsylvania 19104, USA. ⁶⁸Laboratory of Molecular Medicine and Biotechnology, Don C. Gnocchi Foundation IRCCS, S. Maria Nascente, 20148 Milan, Italy. ⁶⁹Department of Medical Epidemiology and Biostatistics, Karolinska Institute, 17177 Stockholm, Sweden. ⁷⁰Atherosclerosis Research Unit, Department of Medicine Solna, Karolinska Institutet, Center for Molecular Medicine, L8:03, Karolinska University Hospital Solna, S-171 76 Stockholm, Sweden. ⁷¹Department of Neurology,

Heinrich-Heine-University, D-40225, Düsseldorf, Germany. ⁷²Commissariat à l'énergie atomique, Institut de Génomique, Centre National de Genotypage, 2 rue Gaston Cremieux, CP 5721, 91057 Evry Cedex, France. ⁷³Experimental and Clinical Research Center, Charité, Universitätsmedizin Berlin and Max Delbrueck Center for Molecular Medicine, Berlin 13125, Germany. ⁷⁴Department for Paediatric Pneumology, Allergy and Neonatology, Hannover Medical School, Carl-Neuberg Strasse 1, D30625 Hannover, Germany. ⁷⁵Centre for Neuroscience, University of Melbourne, Victoria 3010, Australia. ⁶Institute of Epidemiology, Helmholtz Zentrum München, German Research Center for Environmental Health, Ingolstädter Landstrasse 1,85764 Neuherberg, Munich, Germany. ⁷⁷Seinäjoki Central Hospital, Seinäjoki, 60220, Finland. ⁷⁸Hunter Medical Research Institute, John Hunter Hospital, Lookout Road, New Lambton, New South Wales 2305, Australia. ⁷⁹SCDU Neurology, Maggiore della Carità Hospital, 28100 Novara, Italy. ⁸⁰Unit of Public Health Genomics, National Institute for Health and Welfare, Helsinki 00290, Finland. ⁸¹Molecular Medicine and Science for Life Laboratory, Department of Medical Sciences, Uppsala University, Entrance 70, 3rd Floor, Res Dept 2, University Hospital, S-75185 Uppsala, Sweden. ⁸²Human Genetics and Cancer Biology, Genome Institute of Singapore, Singapore 138672⁸³Institute of Immunology, Oslo University Hospital, N-0027 Oslo, Norway. ⁸⁴Institute of Experimental Neurology (INSPE), San Raffaele Scientific Institute, Via Olgettina 58, 20132 Milan, Italy. ⁸⁵Department of Psychiatry and Human Behavior, University of California, Irvine (UCI), 5251 California Av, Ste 240, Irvine, California 92617, USA. ⁸⁶Christchurch School of Medicine, University of Otago, Christchurch 8041, New Zealand. ⁸⁷John P. Hussman Institute for Human Genomics and The Dr. John T Macdonald Foundation Department of Human Genetics, University of Miami, Miller School of Medicine, 1501 NW 10th Avenue, Miami, Florida 33136, USA. ⁸⁸Greater Manchester Centre for Clinical Neurosciences, Hope Hospital, Salford M6 8HD, UK. ⁸⁹The Department of Neurology, Dunedin Public Hospital, Otago 9016, New Zealand. ⁹⁰The Norwegian Multiple Sclerosis Competence Centre, Department of Neurology, Haukeland University Hospital, N-5021 Bergen, Norway. ⁹¹Department of Clinical Medicine, University of Bergen, N-5021 Bergen, Norway. ⁹²Plymouth Hospitals NHS Trust, Department of Neurology, Derriford Hospital, Plymouth PL6 8DH, UK. ⁹³Department of Medical Genetics, University of Helsinki and University Central Hospital, Helsinki 00014, Finland. ⁹⁴Program in Medical and Population Genetics and Genetic Analysis Platform, The Broad Institute of MIT and Harvard, Cambridge, Massachusetts 02142, USA. ⁹⁵Pôle Neurosciences Cliniques, Service de Neurologie, Hôpital de la Timone, 13005 Marseille, France. ⁹⁶Department Neurology, Oulu University Hospital, Oulu 90029, Finland. ⁹⁷UK MS Tissue Bank, Wolfson Neuroscience Laboratories, Imperial College London, Hammersmith Hospital, London W12 0NN, UK. ⁹⁸Université de Montréal & Montreal Heart Institute, Research Center, 5000 rue Belanger, Montreal, Quebec H1T 1C8, Canada. ⁹⁹Neurology and Center for Experimental Neurological Therapy (CENTERS), Sapienza University of Rome, 00189-Rome, Italy. ¹⁰⁰KOS Genetic Srl, Via Podgora, 7, 20123 Milan, Italy. ¹⁰¹Department of General Internal Medicine, University Hospital, Schleswig-Holstein, Christian-Albrechts-University, Kiel 24105, Germany. Systems Biology and Protein-Protein Interaction, Center for Molecular Neurobiology, ¹⁰³ Falkenried 94, D-20251 Hamburg, Germany. ¹⁰³Center for Human Genetics Research, Vanderbilt University Medical Center, 519 Light Hall, Nashville, Tennessee 37232, USA. ¹⁰⁴Telethon Institute for Child Health Research, Centre for Child Health Research, University of Western Australia, 100 Roberts Road, Subiaco, Western Australia 6008, Australia. ¹⁰⁵Cambridge Institute for Medical Research, University of Cambridge School of Clinical Medicine, Cambridge CB2 0XY, UK. ¹⁰⁶Department of Neurology, Helsinki University Central Hospital and Molecular Neurology Programme, Biomedicum, University of Helsinki, FIN-00290 Helsinki, Finland. ¹⁰⁷Division of Psychological Medicine and Psychiatry, Biomedical Research Centre for Mental Health at the Institute of Psychiatry, King's College London and The South London and Maudsley NHS Foundation Trust, Denmark Hill, London SE5 8AF, UK. ¹⁰⁸Service de Neurologie et Faculté de Médecine de Reims, Université de Reims Champagne-Ardenne, 51100 Reims, France. ¹⁰⁹University of Queensland Diamantina Institute, Princess Alexandra Hospital, Brisbane, Queensland 4102, Australia. ¹¹⁰Department of Epidemiology and Population Health, London School of Hygiene and Tropical Medicine, London WCIE 7HT, UK.¹¹¹St. Vincent's University Hospital, Dublin 4, Ireland.¹¹²Neuropsychiatric Genetics Research Group, Institute of Molecular Medicine, Trinity College Dublin, Dublin 2, Eire. ¹¹³Centre for Gastroenterology, Bart's and the London School of Medicine and Dentistry, London E1 2AT, UK.¹¹⁴Department of Neurosciences, Institute of Biomedical Research August Pi Sunyer (IDIBAPS), Hospital Clinic of Barcelona, 08036, Spain.¹¹⁵Clinical Neurosciences, St George's University of London, London SW17 ORE, UK.¹¹⁶Department of Medical and Molecular Genetics, King's College London School of Medicine, Guy's Hospital, London SE1 9RT, UK. ¹¹⁷Medical Research Council Biostatistics Unit, Robinson Way, Cambridge CB2 OSR, UK.¹¹⁸Biomedical Research Institute, University of Dundee, Ninewells Hospital and Medical School, Dundee DD1 9SY, UK. ¹¹⁹Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, 81377 Munich, Germany. ¹²⁰Klinikum Grosshadern, Munich 81377, Germany. ¹²¹King's College London, Social, ¹²⁰Klinikum Grosshadern, Munich 81377, Germany. ¹²¹King's College London, Social, Genetic and Developmental Psychiatry Centre, Institute of Psychiatry, Denmark Hill, London SE5 8AF, UK. ¹²²Department of Neurology, Auckland City Hospital, Grafton Road, Auckland 1010, New Zealand. ¹²³Institut für Humangenetik, Technische Universität München, 81675 Munich, Germany. ¹²⁴Institut für Humangenetik, Helmholtz Zentrum München, 85764 Neuherberg, Munich, Germany. ¹²⁵Popgen Biobank, Christian-Albrechts University Kiel, Kiel 24105, Germany. ¹²⁶Pöle Recherche et Santé Publique, CHU Pontchaillou, 35033 Rennes, France. ¹²⁷NIHR Biomedical Research Centre for Onbthalmology. Moorfields Eve Hospital NHS Foundation Trust and UICI Centre for Ophthalmology, Moorfields Eye Hospital NHS Foundation Trust and UCL Institute of Ophthalmology, London EC1V 2PD, UK. ¹²⁸Department of Molecular Neuroscience, Institute of Neurology, Queen Square, London WC1N 3BG, UK. ¹²⁹Molecular and Physiological Sciences, The Wellcome Trust, London NW1 2BE, UK. ¹³⁰Harvard NeuroDiscovery Center, Harvard Medical School, Boston, Massachusetts 02115, USA. 131 Department of Neurology & Immunology, Yale University Medical School, New Haven, 06520 Connecticut, USA.

*These authors contributed equally to this work.

†Deceased.